Grape Leaffolder, *Desmia funeralis* (Hübner) (Insecta: Lepidoptera: Crambidae)¹

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The Featured Creatures collection provides in-depth profiles of insects, nematodes, arachnids, and other organisms relevant to Florida. These profiles are intended for the use of interested laypersons with some knowledge of biology as well as academic audiences.

Introduction

The grape leaffolder is a common and widely distributed species that is a minor pest of grapes in the United States. In California severe outbreaks have occurred in restricted areas during some years. These outbreaks have been associated with failures of parasites to attain their normal levels. In Florida the grape leaffolder has done substantial damage to grape leaves in September and October when growers have discontinued their spray programs after harvest. Excessive defoliation may deplete the food reserves in the vine sufficiently to reduce the size of the next crop.

The ability to control grape pests in Florida is becoming more meaningful with the rebirth of the grape industry. At one time, bunch grape plantings numbered several thousand acres in Florida, then nearly disappeared from the adverse effects of Pierce's disease. Development of several resistant bunch grape varieties by the former University of Florida Central Florida Research and Education Center at Leesburg spurred renewed interest. In 2017, the combined commercial muscadine and bunch grape acreage in Florida was estimated to be about 1,600 acres (USDA Census of Agriculture, 2017). There are currently 25 farm wineries operating across the state (https://www.fgga.org/florida-winery-directory/). Grapes are also sold as fresh fruit and juice.

Identification

Eggs are small (about ¹/₃₂ inch long), flat, iridescent, elliptical structures laid singly on the underside of a leaf, often in the angles between a vein and the leaf surface.

Larvae are ³/₄ inch long when fully grown. They are glossy, translucent yellow-green on the sides and somewhat darker above, with scattered fine yellow hairs on each segment. The head and prothoracic shield are light brown, and there are light brown spots on the sides of the first two thoracic segments. Larvae wiggle vigorously when disturbed and drop to the ground.

Pupae average a little over ½ inch long; they are light brown just after pupation, but soon turn dark.

Adults have a wing expansion varying from about ⁴/₅ inch to nearly 1¹/₄ inches. The color of the wings is very dark brown, almost black, with a silvery or bluish iridescence. The forewings in both sexes have two nearly oval white spots. The hind wings of the male have one white bar which in the female is partly or completely divided into two spots. Both

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sexes have various amounts of white on the fringes of the wings and parts of the head, body, and legs. The antennae in the male appear thickened and notched near the middle, while in the female they are uniform and threadlike. Adults of other species of *Desmia* and certain other similar species in Florida have the white spots of the wings much more linear or are obviously different in number of spots or some other characteristic.



Figure 1. Larva of the grape leaffolder, *Desmia funeralis* (Hübner). Credits: F. L. Jensen, Farm Advisor, California



Figure 2. Adult grape leaffolder, *Desmia funeralis* (Hübner). Credits: Division of Plant Industry

Life History

Winter is passed in the pupal stage in the folded and fallen leaves. The moths emerge in spring shortly after grape foliage appears and lay their eggs on the leaves. In Florida the earliest records for adults are in mid-February in the Sarasota area, but other February records include localities in Indian River, Manatee, Osceola, and Volusia counties. Adults have been taken all months of the year in Florida except January. Comparatively few larval records are available for Florida, but the earliest one is in May and the latest in December, with peak populations occurring from July into October. The numbers of first brood larvae are reported to be quite insignificant compared to the second brood. In the southern United States a third brood leads into the overwintering pupal stage. Precise data on the developmental stages of Desmia funeralis in Florida are lacking, but Smith and Stafford (1955) give data for Fresno, California that may approximate development in Florida. The moth flight period of the first brood occurred (at Fresno) from April 2 to May 24; egg hatching required 10 to 17 days. Total time in larval stages was three to four weeks, time in the pupal stage was 10 to 14 days, with the total time from egg to emergence of moth 6 1/2 to 7 ¹/₂ weeks. The second brood moth flight period was from June 15 to July 15. Egg hatching required four to five days. Total time in larval stages was two to three weeks, time in the pupal stage seven to 11 days, and total time from egg to emergence of the moth four to five weeks. The third brood moth flight period extended from August 3 to September 5; eggs hatched in four to five days, larvae required three to five weeks to complete growth, and ensuing pupae overwintered. Female moths of the second and third broods lay most of their eggs on leaves rolled by larvae of a previous brood. However, moths may fly into an uninfested vineyard as late as mid-summer and start what may become a serious leaffolder problem.

AliNiazee (1974) studied leaffolder development in the laboratory at $23.9 \pm 1^{\circ}$ C. He found the average duration of larval stages to be as follows: first instar, 3.5 days; second instar, 3.9 days; third instar, 4.4 days; fourth instar, 4.3 days; fifth instar, 6.1 days.

Leaffolders feed on both muscadine and bunch grape leaves. Young larvae will tie together several muscadine leaves and feed in a group. Individual larvae begin making leaf rolls after one week (AliNiazee 1974). They may also use rolls made by an earlier generation. Larvae will fold, rather than roll, bunch grape leaves, which are thicker than muscadine leaves (McGiffen and Neunzig 1985).

Larvae make leaf rolls by spinning strands of silk, which contract and pull the leaf together. Jensen (1966) noted that each bunch of strands contains 200 to 300 individual filaments and about 10 such ties are needed per roll, including some inside where they cannot be seen. Larvae feed on the free edge of the leaf inside the roll and make at least two such rolls during their development. They always remain inside the rolls or between leaves except when moving (at night) from one location to another. To form pupation sites, larvae make envelopes, often by cutting a small piece of leaf on three sides and folding it over (AliNiazee 1974). At the end of the season these will fall to the ground with the leaves to which they are attached. AliNiazee (1974) monitored adult activity in the summer in California and found that moths were most active from midnight to the early morning hours.

Detection and Damage

Injury to the leaves is very characteristic and may be easily recognized. As soon as a larva is large enough it folds the leaf, exposing the under surface; the edge is held in place by bands of silk thread. It is within the protection of this fold that the larva feeds, skeletonizing the leaf of the upper surface. When the larvae are numerous, the injury to the vine becomes conspicuous, even at a considerable distance, because the light color of the under surface of the folded leaves contrasts boldly with the dark green of the upper side normally presented, thus giving the vine a patchy appearance. Larvae roll muscadine leaves, which are thinner than bunch grape leaves.



Figure 3. Damage to bunch grape foliage caused by the grape leaffolder, *Desmia funeralis* (Hübner). Credits: Warren Adlerz, UF/IFAS

Hosts

Wild and cultivated grapes, *Vitis* spp., are the principal hosts of the grape leaffolder. Two varieties of redbud, *Cercis canadensis* and *C. chinensis*; Virginia creeper, *Parthenocissus quinquefolia*; and *Oenothera* have been reported as occasional hosts. The varieties of grape more prone to attack are those having thinner, more tender, and hairy leaves.

Parasitoids

Many parasitoids have been reported from grape leaffolder. McGiffen and Neunzig (1985) reared larvae and found three species of tachinid flies as well as numerous hymenopterous parasitoids. In all, they found three species of braconid wasps, two species of ichneumonid wasps, and one eulophid wasp.



Figure 4. Damage to muscadine grape leaf with characteristic roll caused by the grape leaffolder, *Desmia funeralis* (Hübner), also showing silk strands. Larva removed from rolls for photographic purposes.

Credits: F.L. Jensen, Farm Advisor, California

Management

Effective insecticides include methoxyfenozide, chlorantraniliprole (rynaxypyr), Spinosad, and spinetoram. Double-check state recommendations before applying any insecticide. Tests in California and Florida using preparations of *Bacillus thuringiensis* (Bt) showed no significant differences in control between it and carbaryl, with both providing good control. Home gardeners who do not want to use insecticides may get good results by thoroughly going over vines when folded or rolled leaves are in evidence and crushing the larvae by hand. Because the insect passes the winter in the pupal stage in fallen leaves, it is a good idea to rake up the leaves and destroy them.

Selected References

AliNiazee MT. 1974. Contribution to the bionomics of the grape leaffolder, *Desmia funeralis* (Hübner): A laboratory study with field observations. Pan-Pacific Entomol. 50: 269–278.

Jensen FL. 1966. Grape leaffolder control with *Bacillus thuringiensis*. Calif. Agric. 20: 2–3.

Kimball CP. 1965. The Lepidoptera of Florida; an annotated checklist. Arthropods of Florida and neighboring land areas Vol. 1. FL Dep. Agric., Div. Plant Industry. 363 p.

McGiffen KC., Neunzig HH. 1985. A guide to the identification and biology of insects feeding on muscadine and bunch grapes in North Carolina. NC Agric. Res. Serv. Bull. 470. 93 pp.

Smith LM, Stafford EM. 1955. Grape pests in California. CA Agric. Exp. Sta. Circ. 445: 16–19.

Stafford EM, Jensen FL. 1957. Grape leaffolder: field tests compared effectiveness of insecticides in control of vineyard pest. Calif. Agric. 11: 4, 8, 10, 15.

Strauss JF. 1916. The grape leaf-folder. USDA Bull. 419: 1–14.

UC IPM. 2016. UC IPM Pest Management Guidelines: Grape: Grape Leaffolder. UC ANR Publication 3448. https://www.ipm.ucanr.edu/agriculture/grape/grapeleaffolder.

USDA Census of Agriculture. 2017. https://www.nass.usda. gov/Publications/AgCensus/2017/Full_Report/Volume_1,_ Chapter_1_State_Level/Florida/st12_1_0037_0037.pdf